1	Supplementary Information to
2	FELASA Working Group Report
3	Capture and Transport of live cephalopods:
4	recommendations for scientific purposes
5	
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# 61 Reference to the Ancillary work

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## 70 Legislative framework

We analyzed the available legislation and recommendations regarding capture and transport of aquatic animals, and identified more than 20 different documents<sup>a</sup>. Here we do not provide a detailed overview about the legislative framework, but refer to the ancillary work (see Pieroni et al., 2021) and suggest the Reader to refer to it for completeness (see also table 1 in Pieroni et al., 2021).

Our analysis revealed that most of the recommendations are aimed at protecting animals during transportation, with limited detail provided for the capture from the wild, at least when aquatic species are considered. Another key- and reiterated element in these norms is the attention to the competence of personnel; i.e., the need for a proper training for those involved in the capture and transport of wild animals. All of them point out that the main source of suffering and distress in animals being

<sup>&</sup>lt;sup>a</sup> We refer to: 86/609/EEC; 91/67/EEC; ETS No.193; EC No 1/2005; 2006 No. 3260; 2007/526/EC; Directive 2010/63/EU; EAZA Position Statement on Council Regulation 1/2005: Protection of Animals during Transport; NC3Rs' best practice for animal transport; Guidelines for the Transport of Laboratory Animals; Conf. 10.21 (Rev. CoP16); Packer's Guidelines Inv1/Aquatic invertebrates; Aquatic Animal Health Code; Guide for the Care and Use of Laboratory Animals; Australian code for the care and use of animals for scientific purposes 8th Edition 2013; Olfert et al., 1993.

captured, handled, and transported is related to the limited competence of peopleinvolved.

Available legislation and guidelines also include information about proper *i*. documentation and planning of the journey, *ii*. methods and means of transport including design of containers, *iii*. health assessment, food, and water supplies for the animals, *iv*. the need of acclimatization before and after transport of wild animals. All the documents we considered have very little information (or any) relevant to cephalopod molluscs.

90

# 91 Competence and attitude of the personnel involved in capture and92 transportation

93 The team of collectors or fishers and shippers has to be properly trained for the scope, 94 and it is responsibility of Authorities to verify that this will occur. Depending on the 95 cases, the team should avail of the support and advice of a Designated Veterinarian 96 who shall check the health status of the animals and shall take the proper actions to 97 avoid or end suffering.

Annex IV of the EU Council Regulation No 1/2005<sup>1</sup> states that drivers and attendants 98 (as referred to in Article 6(5) and Article 17(1)) shall have successfully completed the 99 100 training and have passed an examination approved by the NCA, ensuring that 101 examiners are independent. The courses should include technical and administrative 102 aspects of Community legislation concerning the protection of animals during transport, including animal physiology (e.g., drinking and feeding needs), animal 103 behavior and the concept of stress. Moreover, practical aspects of animal handling and 104 of the emergency care both for the personnel and the animals should be considered in 105 106 the training program assuring that personnel involved (drivers) should know how the driving behavior might impact on the welfare of the transported animals thus also
impacting the quality of meat, if the animals are destined for food consumption (see
Annex IV)<sup>1</sup>.

110

## 111 Principles for Transport, documentation and planning of the journey

Regardless of the purpose of capture and transport of animals, the main principles included in the guidelines and legislations (review in Pieroni et al., 2021) concerning the planning phases prior collection and transport, may be applied to cephalopods after the due considerations.

116 Transport, and in particular long journeys, might be detrimental for the health 117 conditions of animals<sup>1, 2</sup> thus, all the planning should be done in advance and the 118 transport should be carried out without delay to the place of destination.

According to the available knowledge that with increased duration of confinement for
transport, both consumption of available oxygen and detrimental changes in water
chemistry (e.g., accumulation of ammonia, carbon dioxide and depletion of oxygen),
we can distinguish<sup>3</sup>:

i. Short duration journey (< 2h; e.g., from local capturing site to local establishment).</li>
Although this is most likely the less stressful situation, there are important
conditions that should be monitored before and during the trip to the final location
(e.g., pH, temperature and salinity, saturation of oxygen, light exposure).
Vibration, noise, and any other kind of interference should be limited and/or
avoided; attention to the infliction of any physical damage should be constantly
given.

130 ii. Long duration journey (> 2h; e.g., between towns, countries and/or
131 intercontinental). The same precautions described for the short duration trip
132 applies also to this situation. However, considering that the journey might imply
133 the shipping of the animals by sea or by air, special attention should be given to
134 the containers used for this kind of transportations. Care should be provided to

include the documentation concerning management of the welfare state of the
animals during and after the journey (see Supplementary Info and Pieroni et al.,
2021).

138

During a short-duration transport, plastic bags may be replaced with large plastic 139 buckets or boxes with a lid containing sufficient pre-oxygenated seawater to allow the 140 animal to be completely immersed<sup>3, 4</sup>. For long-duration transport and similarly to 141 fish<sup>5</sup>, considering animals' body size, the cephalopod should be placed with adequate 142 143 volume of seawater and oxygen-enriched air in double common aquarium aerated bags (see Table 2). For transport periods over 12 hours, aeration and oxygenation may 144 be necessary being careful not to induce distress to the animals (i.e., water turbulence 145 and bubbling can cause air entrapment in the mantle cavity or produce microbubbles 146 affecting the integrity of the mucus layer on the skin). Sealed holding bags containing 147 oxygenated seawater should be placed into insulated boxes (e.g., Styrofoam) to ensure 148 that a temperature, appropriate to the species, is maintained during transport<sup>3</sup>. Bags 149 should be packed with cushioning material (e.g., paper, Styrofoam pellets) to ensure 150 151 they do not move during transport and the external shipping box should report the labels: "this side up" and "live animals" (see 'Other General Requirements' in Table 152 2). 153

154

In Europe, but also according to other national legislations (see table 1 in Pieroni et al., 2021), every transportation should be preceded by accurate planning of the journey and shall be accompanied by a proper documentation stating: **a**) the origin and their ownership; **b**) the place of departure; **c**) the date and time of departure; **d**) the intended

place of destination; e) the expected duration of the intended journey (see Chapter II, 159 Art. 4 and Appendix included in EU Council Regulation No 1/2005)<sup>1</sup>. The regulation 160 recommend that a competent scientific committee shall be consulted about the 161 162 duration and the route plans of animal transport, and according to a harmonized European model, certificate for transporters must be presented. Authorities should 163 164 shall take the necessary measures to prevent or reduce to a minimum any delay during transport (e.g., programming special arrangements at the place of transfers, exit points 165 and border inspections to give priority to the transport of animals) but resting periods 166 at specific control posts shall be planned if the journey is longer (Art. 22(1))<sup>1</sup>. 167 Furthermore, the consignment shall be done immediately unless the detainment is 168 necessary for the health of the animal or for public safety, such as possible spread of 169 zoonosis if there are some diseased animals (Art. 22(2))<sup>1</sup>. For such a reason, veterinary 170 checks at borders' inspection include the analysis of the welfare conditions in which 171 the animals are transported. 172

Further documentation is needed if transporting wild, timid or dangerous species,
providing instructions about feeding, watering and any special care required for them
(see Chapter II, Section 1.3)<sup>1</sup>.

The person planning the journey has to bring and compile a journey log, which reports in detail any daily event including animals' health status, any intervention performed and any detour from the original plan (see Annex II)<sup>1</sup>. As for the regulation of the international transport of animals the ETS No.193<sup>6</sup> applies.

180

On the basis of EU legislation and other documents - such as ETS No.193 and Council Recommendation 2007/526/EC (see Pieroni et al., 2021 for details) - specifically addressed to experimental animals, LASA produced a WG Report 'Guidance on the transport of laboratory animals'<sup>7</sup> which lists all the documentation needed during the planning of the journey (see Paragraph 3.3<sup>7</sup>). Of course, the number and type of documents to be filled up depend on the journey type, species, microbiological status,and route. However, the following information should not be missing:

- *i.* shipment documentation details such as waybill number or IATA Shipper's certificate (for Air transport), import licenses issued by the State Veterinary Service, CITES permits where necessary (for intra-European and Third-country shipping), invoices for Customs purposes, health certificate of the animal transported signed by the Designated Veterinarian, journey log or transfer authorizations from specific bodies that regulate laboratory animals use;
- *ii.* animal details such as species, strain, scientific name, number, sex, age,
  weight, identification numbers or any special requirements resulting from
  phenotype;
- *iii.* personnel details such as contact information of sender, intermediaries,
  consignee, shipper/carrier, veterinarian;
- *iv.* crates with date and times the animals were packed loaded, and departed
  with clear 'Live animals' and orientation arrows labels; *v.* expected events,
  such as proposed and actual rest periods, pre-journey review of plan by
  consignor and post-journey review of plan by new owner<sup>7</sup> (see also Pieroni
  et al., 2021 for details).

205

# 206 Different forms, different methods: considerations for Capture and 207 Transport methods of different cephalopods' life stages

- It is essential to take into account the life stage of the target cephalopod species.
- 209 Live cephalopods or eggs?
- 210 Collection and standardized transport of eggs for target cephalopod species have been
- 211 proposed as an alternative, since these appear easier to manage.

For 'classic' laboratory species (e.g., fish), moving of embryos, sperm or eggs between 212 research facilities has increased and is now well established. This approach is also 213 considered as a way to comply with Refinement since transport appear to be stressful 214 for live animals and could impact their welfare, while for early life stages it seems 215 limited. On the other hand, the use of captive animals (raised from eggs) would allow 216 to fulfill experimental requirements, such as controlling previous experiences. Under 217 similar considerations, the use of animals taken from the wild would be in accordance 218 219 with the principle of using animals that have been 'primed' by natural environmental stimuli. 220

221

222 When applied to cephalopods, considerations should be taken to offer a standardized method of egg collection to reduce the impact over natural resources (e.g., collecting 223 224 those stranded on the beach or those captured as by-catch), and to minimize the harm to the egg masses. Though, cephalopods eggs require delicacy in handling, accurate 225 temperature and salinity control during all phases, including transportation<sup>8, 9</sup>. It is 226 227 noteworthy to remind that eggs collected at advanced stages of development may induce premature hatch during transport and because of handling as they also have 228 higher metabolic requirements<sup>10</sup>, and - for some cephalopod species - maternal care is 229 needed in order to develop in a proper and healthy way. 230

231 Methods for transport of eggs of several cephalopod species have been developed 232 achieving some standardization. Examples are available for cuttlefish, squid and 233 octopus eggs<sup>9, 11</sup>.

The Directive 2010/63/EU considers the protection of this taxon from hatching. 235 However, several studies are based on the collection and culture of egg masses from 236 the wild which - although easier to obtain and then transport in terms of size of 237 containers and water volume - require particular care, being very sensitive to even 238 little changes in temperature, pH and salinity (for example see Iglesias et al., 2007)<sup>12</sup>. 239 Storing conditions during embryonic development should also be monitored because 240 eggs health state will consequently affect the "quality" of the hatchlings. As 241 mentioned, it is also crucial to consider whether maternal care is required, like in 242 incirrate octopods or oceanic squids<sup>13, 14</sup> and attempts to properly simulate are 243 mandatory. Maternal care is a critical factor for the proper embryonic development 244 and to limit the risk of premature hatching<sup>15-19</sup>. 245

246

#### 247 From hatchlings to adult

Cephalopod hatchlings are either miniature adults or planktonic paralarvae with 248 relatively short arms and limited swimming ability<sup>20</sup> and represent extremely delicate 249 developing forms, very sensitive to any insult or change in the water parameters from 250 251 the site of collection to the containers. Nevertheless, the majority of studies (see table 2 in Pieroni et al., 2021) used trawls and bongo nets for collection, with few reporting 252 also that animals which resulted damaged were excluded from the experiments (for 253 example see Otero et al., 2016)<sup>21</sup>. A few notes about capture and transport of paralarval 254 stages of cephalopods is given in the ancillary work to this paper (Pieroni et al., 2021). 255

The collection of juveniles is a challenging task because these may be selective and with rigid feeding requirements<sup>22</sup>; appropriate cautions should be applied in these circumstances, because a traumatic capture might affect highly feeding and behavioral responses that may translate into fatal conditions.

261

Adults have also important temperature, salinity and pH requirements as most of the cephalopods are stenotherm and stenohaline. It follows that the inappropriate capture and transport of any stage from the wild might result in high mortality and therefore considerable species-specific care is needed if viable animals have to be returned to the laboratory<sup>3, 11, 23</sup>. Further information and summary of requirements are provided in Pieroni et al. (2021).

268

## 269 Other Recommendations for live cephalopods transport

Prior to transport, and before long-journeys, wild-caught cephalopods should be
acclimated to captivity in aquaria and should be monitored during this phase,
checking for good appetite, presence of any skin lesions and potential unusual
behaviour.

274 If the journey to the lab is brief, small sepioids and octopuses could be temporarily 275 placed in containers part-filled with seawater. If temperature, pH and oxygen content 276 values change, renewal of seawater is mandatory<sup>4</sup>. For transport lasting more than one or two hours, small numbers of small individuals could be carried in cooled boxes
with only sufficient water to cover the animals, each one contained in a polythene bag
about 1/3 filled with seawater with oxygen filling the remaining space. Survival for 810 h was reported to be easily possible by sealing and keeping the bags cool.

281

Another factor to consider is food deprivation. It is suggested to prevent animals from 282 feeding for 24 hours before long-term journeys because it helps keeping metabolic rate 283 284 under baseline, possibly limiting ammonia build-up during transport. However, food deprivation depends on the animal normal feeding frequency, oro-anal transit time 285 and renal ammonium ion excretion for the species<sup>24</sup>. Sedation is not essential and is 286 287 not recommended for transport of most cephalopods. However, sedation methods have been utilized for the transport of some species, in some cases with controversial 288 289 results. Interestingly Grimpe<sup>11</sup> suggested that very long duration transportation - i.e. requiring more than two days - should be achieved in steps allowing 'resting' periods 290 291 in appropriate locations which is what nowadays is indicated by the European and 292 international legislations for the transport of live animals (for details see Pieroni et al., 2021). 293

296	References
297	
298	1. Council of the European Union. Council Regulation (EC) No 1/2005 of 22 December
299	2004 on the protection of animals during transport and related operations and amending
300	Directives 64/432/EEC and 93/119/EC and Regulation (EC) No 1255/97, (2004, accessed
301	https://eur-lex.europa.eu/legal-content/en/ALL/?uri=CELEX%3A32005R0001).
302	2. Commission of the European Communities. Commission Recommendation of 18
303	June 2007 on guidelines for the accommodation and care of animals used for experimental
304	and other scientific purposes (notified under document number C(2007) 2525), (2007,
305	accessed <u>https://eur-lex.europa.eu/legal-content/EN/TXT/?uri=CELEX%3A32007H0526</u> ).
306	3. Fiorito G, Affuso A, Basil J, et al. Guidelines for the Care and Welfare of
307	Cephalopods in Research - A consensus based on an initiative by CephRes, FELASA and the
308	Boyd Group. <i>Lab Anim</i> 2015; 49: 1-90.
309	4. Budelmann BU. Cephalopoda. <i>The UFAW handbook on the care and management of</i>
310	laboratory and other research animals 2010: 787-817.
311	5. Berka R. The Transport of Live Fish: A Review. Rome: European Inland Fisheries
312	Advisory Commission (EIFAC); FAO - Food and Agriculture Organization of the United
313	Nations, 1986.
314 215	b. Council of Europe. European Convention for the Protection of Animals during
316	https://www.coe.int/en/web/conventions/full-list/_/conventions/rms/0900001680083710)
310	7 Swallow I Anderson D Buckwell AC et al Guidance on the transport of laboratory
318	animals. <i>Lab Anim</i> 2005: 39: 1-39. 2005/02/11. DOI: 10.1258/0023677052886493.
319	8. Villanueva R, Vidal EAG, Fernández-Álvarez FÁ, et al. Early Mode of Life and
320	Hatchling Size in Cephalopod Molluscs: Influence on the Species Distributional Ranges.
321	PLOS ONE 2016; 11: e0165334. DOI: 10.1371/journal.pone.0165334.
322	9. Deryckere A, Styfhals R, Vidal EAG, et al. A practical staging atlas to study
323	embryonic development of Octopus vulgaris under controlled laboratory conditions. BMC
324	Developmental Biology 2020; 20: 7. DOI: 10.1186/s12861-020-00212-6.
325	10. Boletzky S. Recent studies on spawning, embryonic development, and hatching in
326	the Cephalopoda. Advances in Marine Biology. Elsevier, 1989, pp.85-115.
327	11. Grimpe G. Pflege, Behandlung und Zucht der Cephalopoden fur zoologische und
328	physiologische Zwecke. In: Äberhalden E (ed) Handbuch der biologischen Arbeitsmethoden.
329	Berlin, Wien: Verlag Urban & Schwarzenberg, 1928, pp.331-402.
330	12. Iglesias J, Sánchez FJ, Bersano JGF, et al. Rearing of Octopus vulgaris paralarvae:
331	present status, bottlenecks and trends. <i>Aquaculture</i> 2007; 266: 1-15.
332	13. Seibel BA, Robison BH and Haddock SHD. Post-spawning egg care by a squid.
333	Nature 2005; 438: 929-929. DOI: 10.1038/438929a.
334	14. Bush SL, Hoving HJT, Huffard CL, et al. Brooding and sperm storage by the deep-
335	sea squid bathyteuthis berryi (Cephalopoda: Decapodiformes). Journal of the Marine Biological
330 227	Association of the United Kingdom 2012; 92: 1629.
337 220	10. Nosa N, r internet ivio, boavida-roriugai J, et al. Ocean Warming Ennances Malformations, Promature Hatching, Metabolic Suppression and Ovidative Stress in the
550	manormations, riemature matching, metabolic suppression and Oxidative stress in the

- Early Life Stages of a Keystone Squid. *PLOS ONE* 2012; 7: e38282. DOI:
- 340 10.1371/journal.pone.0038282.
- 16. Vidal EAG, Villanueva R, Andrade JP, et al. Chapter One Cephalopod Culture:
- 342 Current Status of Main Biological Models and Research Priorities. In: Vidal EAG (ed)
   343 Advances in Marine Biology. Academic Press, 2014, pp.1-98.
- 344 17. Boletzky Sv. Biology of early life stages in cephalopod molluscs. *Advances in Marine*345 *Biology* 2003; 44: 143-203.
- 18. Nabhitabhata J, Asawangkune P, Amornjaruchit S, et al. Tolerance of eggs and
- hatchlings of neritic cephalopods to salinity changes *Phuket Marine Biological Center Special Publication* 2001; 25: 91-99.
- 349 19. Bower JR and Sakurai Y. Laboratory observations on *Todarodes pacificus*
- 350 (Cephalopoda: Ommastrephidae) egg masses. *American Malacological Bulletin* 1996; 13: 65-71.
- Villanueva R. Experimental rearing and growth of planktonic Octopus vulgaris from
  hatching to settlement. *Canadian Journal of Fisheries and Aquatic Sciences* 1995; 52: 2639-2650.
- 352 natching to settlement. *Cunatum Journal of Fisheries and Aquatic Sciences* 1995, 52, 2059-2050.
   353 21. Otero J, Álvarez-Salgado XA, González ÁF, et al. Wind-driven upwelling effects on
- 354 cephalopod paralarvae: Octopus vulgaris and Loliginidae off the Galician coast (NE
- 355 Atlantic). *Progress in Oceanography* 2016; 141: 130-143.
- 35622.Boletzky SV and Hanlon RT. A Review of the Laboratory Maintenance, Rearing and
- Culture of Cephalopod Molluscs. *Memoirs of the National Museum of Victoria* 1983; 44: 147-186.
- **358** 23. Boyle PR. *The UFAW handbook on the care and management of cephalopods in the*
- *laboratory*. Potters BarUniversities Federation for Animal Welfare, 1991, p.915.
- 360 24. Sykes AV, Almansa E, Cooke GM, et al. The digestive tract of cephalopods: a
- neglected topic of relevance to animal welfare in the laboratory and aquaculture. *Frontiers in physiology* 2017; 8: 492.

# 365 Additional References – Overview of Capture and Transport Methods

- **366** List of References in support of Table 1 Capture Methods
- 367
- 368 General
- Andrews PLR, Darmaillacq AS, Dennison N, Gleadall IG, Hawkins P, Messenger JB, Osorio
  D, Smith VJ and Smith JA. The identification and management of pain, suffering and
  distress in cephalopods, including anaesthesia, analgesia and humane killing. *J Exp Mar Biol Ecol* 2013; 447: 46-64.
- Boletzky SV and Hanlon RT. A Review of the Laboratory Maintenance, Rearing and Culture
  of Cephalopod Molluscs. *Memoirs of the National Museum of Victoria* 1983; 44: 147-186.
- Fiorito G, Affuso A, Anderson DB, Basil J, Bonnaud L, Botta G, Cole A, D'Angelo L, de
  Girolamo P, Dennison N, Dickel L, Di Cosmo A, Di Cristo C, Gestal C, Fonseca R, Grasso
  F, Kristiansen T, Kuba M, Maffucci F, Manciocco A, Mark FK, Melillo D, Osorio D, Palumbo
- A, Perkins K, Ponte G, Raspa M, Shashar N, Smith J, Smith D, Sykes A, Villanueva R,
   Tublitz N, Zullo L and Andrews PLR. Cephalopods in neuroscience: Regulations, Research
- and the 3Rs. *Invert Neurosci* 2014; 14: 13-36.
- Fiorito G, Affuso A, Basil J, Cole A, de Girolamo P, D'Angelo L, Dickel L, Gestal C, Grasso F,
  Kuba M, Mark F, Melillo D, Osorio D, Perkins K, Ponte G, Shashar N, Smith D, Smith J and
  Andrews PL. Guidelines for the Care and Welfare of Cephalopods in Research A
  consensus based on an initiative by CephRes, FELASA and the Boyd Group. *Lab Anim* 2015;
  49: 1-90.
- Ponte G, Andrews P, Galligioni V, Pereira J and Fiorito G. Cephalopod welfare, biological and
   regulatory aspects: an EU experience. *The Welfare of Invertebrate Animals*. Springer, 2019,
   pp.209-228.
- Rathjen WF. Cephalopod capture methods: an overview. *Bulletin of Marine Science* 1991; 49:
  494-505.
- 391

## 392 Nautilus

- 393 Carlson BA. Collection and aquarium maintenance of Nautilus. In: N.H SWB and Landman394 (eds) *Nautilus*. Springer, 2010, 1991, pp.563-578.
- Bunstan A, Bradshaw CJA and Marshall J. Nautilus at risk–estimating population size and
   demography of *Nautilus pompilius*. *PloS one* 2011; 6: e16716.
- Linzmeier BJ, Kozdon R, Peters SE and Valley JW. Oxygen isotope variability within *Nautilus*shell growth bands. *PLoS One* 2016; 11: e0153890.
- Muntz WRA. Effects of light on the efficacy of traps for *Nautilus pompilius*. *Marine Behaviour and Physiology* 1994; 24: 189-193.
- 401 Oba T, Kai M and Tanabe K. Early life history and habitat of *Nautilus pompilius* inferred from
  402 oxygen isotope examinations. *Marine Biology* 1992; 113: 211-217.
- 403 O'Dor RK, Wells J and Wells MJ. Speed, jet pressure and oxygen consumption relationships
  404 in free-swimming Nautilus. *Journal of experimental biology* 1990; 154: 383-396.
- Uchiyama K and Tanabe K. Hatching of *Nautilus Macromphalus* in the Tobaaquarium, Japan.
   *Advancing research on living and fossil cephalopods*. Springer, 1999, pp.13-16.

## 407 Cuttlefish

- 408 Deepak S and Patterson J. Experimental culture of the Pharaoh's cuttlefish *Sepia pharaonis*,
  409 Ehrenberg 1831, under closed circulation systems. *International Journal of Molecular Sciences*410 2011; 40: 841-846.
- Moltschaniwskyj NA, Hall K, Lipinski MR, Marian JEAR, Nishiguchi M, Sakai M, Shulman
  DJ, Sinclair B, Sinn DL, Staudinger M, Van Gelderen R, Villanueva R and Warnke K. Ethical
  and welfare considerations when using cephalopods as experimental animals. *Reviews in*
- 414 *Fish Biology and Fisheries* 2007; 17: 455-476.
- 415 O'Brien CE, Bellanger C, Jozet-Alves C, Mezrai N, Darmaillacq A-S and Dickel L. Stressful
  416 conditions affect reproducing cuttlefish (*Sepia officinalis*), reducing egg output and quality.
  417 *ICES Journal of Marine Science* 2018; 75: 2060-2069.
- Pereira F, Vasconcelos P, Moreno A and Gaspar MB. Catches of *Sepia officinalis* in the smallscale cuttlefish trap fishery off the Algarve coast (southern Portugal). *Fisheries Research*2019; 214: 117-125.
- 421 Şen H. Effects of String diameter preference of *Sepia officinalis* (L. 1758) during spawning in
  422 captivity. *Journal of Fisheries Sciences* 2013; 7: 297-301.
- Solé M, Monge M, André M and Quero C. A proteomic analysis of the statocyst endolymph
  in common cuttlefish (*Sepia officinalis*): an assessment of acoustic trauma after exposure to
  sound. *Scientific reports* 2019; 9: 1-12.
- Watanuki N and Kawamura G. A review of cuttlefish basket trap fishery. *South Pacific Study*1999; 19: 31-48.
- Watanuki N, Hirayama I and Kawamura G. Why do cuttlefish *Sepia esculenta* enter basket
  traps? Space occupation habit hypothesis. *Fisheries science* 2000; 66: 190-197.
- 430
- 431 Sepiolidae
- 432 Montgomery MK and McFall-Ngai M. Embryonic development of the light organ of the
  433 sepiolid squid *Euprymna scolopes* Berry. *The Biological Bulletin* 1993; 184: 296-308.
- 434 Nabhitabhata J and Nishiguchi MK. *Euprymna hyllebergi* and *Euprymna tasmanica*. *Cephalopod* 435 *Culture*. Springer, 2014, pp.253-269.
- Wei SL and Young RE. Development of symbiotic bacterial bioluminescence in a nearshore
  cephalopod, *Euprymna scolopes*. *Marine Biology* 1989; 103: 541-546.
- 438

### 439 Squid

- Balch N, O'Dor R and Helm P. Laboratory rearing of rhynchoteuthions of the ommastrephid
  squid *Illex illecebrosus* (Mollusca: Cephalopoda). *Vie et Milieu/Life & Environment* 1985: 243246.
- Bower JR, Sakurai Y, Yamamoto J and Ishii H. Transport of the ommastrephid squid *Todarodes pacificus* under cold-water anesthesia. *Aquaculture* 1999; 170: 127-130.
- Budelmann BU. Cephalopoda. *The UFAW handbook on the care and management of laboratory and other research animals* 2010: 787-817.
- Buresch KC, Maxwell MR, Cox MR and Hanlon RT. Temporal dynamics of mating and
  paternity in the squid *Loligo pealeii*. *Marine Ecology Progress Series* 2009; 387: 197-203.

- Cabanellas-Reboredo M, Alós J, Palmer M, Grädel R and Morales-Nin B. Simulating the
  indirect handline jigging effects on the European squid *Loligo vulgaris* in captivity. *Fisheries research* 2011; 110: 435-440.
- 452 Dawe EG, O'Dor RK, Odense PH and Hurley GV. Validation and Application of an Ageing
  453 Technique for Short-finned Squid (*Illex illecebrosus*). *Journal of Northwest Atlantic Fisheries*454 *Science* 1985; 6: 107-116.
- 455 Durholtz MD and Lipinski MR. Influence of temperature on the microstructure of statoliths
  456 of the thumbstall squid *Lolliguncula brevis*. *Marine Biology* 2000; 136: 1029-1037.
- Flores EEC, Igarashi S, Mikami T and Kobayashi K. Studies on Squid Behavior in Relation to
  Fishing: I. On the handling of squid, *Todarodes pacificus* Steenstrup, for behavioral study. *Bulletin of the Faculty of Fisheries, Hokkaido University*. 1976; 27: 145-151.
- Gonçalves JM, Porteiro FM, Cardigos F, Martins HR and Pham CK. The Azorean *Loligo forbesi*(Cephalopoda: Loliginidae) in captivity: transport, handling, maintenance, tagging and
  survival. *Marine Biodiversity Records* 2009; 2: e120.
- Kaplan MB, Mooney TA, McCorkle DC and Cohen AL. Adverse effects of ocean acidification
  on early development of squid (*Doryteuthis pealeii*). *PLoS One* 2013; 8: e63714.
- Matsumoto G. Transportation and maintenance of adult squid (*Doryteuthis bleekeri*) for
   physiological studies. *The Biological Bulletin* 1976; 150: 279-285.
- 467 Perretti CT, Zerofski PJ and Sedarat M. The spawning dynamics of California market squid
  468 (*Doryteuthis opalescens*) as revealed by laboratory observations. *Journal of Molluscan Studies*469 2016; 82: 37-42.
- 470 Puneeta P, Vijai D, Yoo H-K, Matsui H and Sakurai Y. Observations on the spawning behavior,
  471 egg masses and paralarval development of the ommastrephid squid *Todarodes pacificus* in
  472 a laboratory mesocosm. *Journal of Experimental Biology* 2015; 218: 3825-3835.
- 473 Summers WC and McMahon JJ. Studies on the maintenance of adult squid (*Loligo pealei*). I.
  474 Factorial survey. *The Biological Bulletin* 1974; 146: 279-290.
- Thorrold SR. Evaluating the performance of light traps for sampling small fish and squid in
  open waters of the central Great Barrier Reef lagoon. *Marine ecology progress series Oldendorf*1992; 89: 277-285.
- Toyofuku T and Wada T. Chromatophore arrangement and photophore formation in the early
  development of swordtip squid *Uroteuthis (Photololigo) edulis. Fisheries science* 2018; 84: 915.
- Vidal EAG, Villanueva R, Andrade JP, Gleadall IG, Iglesias J, Koueta N, Rosas C, Segawa S,
  Grasse B, Franco-Santos RM, Albertin CB, Caamal-Monsreal C, Chimal ME, EdsingerGonzales E, Gallardo P, Le Pabic C, Pascual C, Roumbedakis K and Wood J. Chapter One
  Cephalopod Culture: Current Status of Main Biological Models and Research Priorities.
- 485 In: Vidal EAG (ed) *Advances in Marine Biology*. Academic Press, 2014, pp.1-98.
- Zakroff C, Mooney TA and Wirth C. Ocean acidification responses in paralarval squid
  swimming behavior using a novel 3D tracking system. *Hydrobiologia* 2018; 808: 83-106.

489 C	)ctopus
-------	---------

- Borges TC, Calixto P and Sendão J. The common octopus fishery in South Portugal: a new
  shelter-pot. *Mediterránea Serie de Estudios Biológico* 2015: 130-154. DOI:
  10.14198/MDTRRA2015.ESP.07.
- Boyle PR. *The UFAW handbook on the care and management of cephalopods in the laboratory*. Potters
  Bar Universities Federation for Animal Welfare, 1991, p.915.
- 495 Carreira GP and Gonçalves JM. Catching *Octopus vulgaris* with traps in the Azores: first trials
  496 employing Japanese baited pots in the Atlantic. *Marine Biodiversity Records* 2009; 2: e114.
- 497 Iglesias J and Fuentes L. *Octopus vulgaris*. Paralarval culture. *Cephalopod culture*. Springer, 2014,
   498 pp.427-450.
- 499 Şen H. Individual rearing of common octopus (*Octopus vulgaris* Cuvier, 1797) in tanks:
  500 Preliminary results. *Ege Journal of Fisheries and Aquatic Sciences* 2019; 36: 361-366.
- 501
- 502

### **503** List of References in support of Table 2 – Transport Methods

- 504
- 505 General
- Aquatic Animals Commission. Control of aquatic animal health risks associated with
  transport of aquatic animals. In: Aquatic Animals Commission (ed) *Aquatic Animal Health Code*. Paris: OIE, World Organization for Animal Health, 2019, pp.4.
- 509 Convention on International Trade in Endangered Species. Packer's Guidelines Inv/1 –
   510 Aquatic invertebrates, https://cites.org/eng/resources/transport/inv1.shtml (1979).

Council of the European Union Council Regulation (EC) No 1/2005 of 22 December 2004 on
the protection of animals during transport and related operations and amending Directives
64/432/EEC and 93/119/EC and Regulation (EC) No 1255/97, (2004, accessed https://eurlex.europa.eu/legal-content/en/ALL/?uri=CELEX%3A32005R0001).

- Gleadall IG. Low dosage of magnesium sulphate as a long-term sedative during transport of
  firefly squid, *Watasenia scintillans*. *Journal of experimental marine biology and ecology* 2013; 447:
  138-139.
- James R. *The use of Anaesthetic and buffer solution in the transportation of cuttlefish-Sepia officinalis.*Internal document. 1992. Weymouth, US: Sea Life Centre Weymouth Biological Services.
- 520
- 521 Nautilus
- 522 Carlson BA. Collection and aquarium maintenance of Nautilus. In: N.H SWB and Landman523 (eds) *Nautilus*. Springer, 2010, 1991, pp.563-578.
- 524
- 525 Cuttlefish
- Jones NJE, Ridgway ID and Richardson CA. Transport of cuttlefish, *Sepia officinalis*, eggs
  under dry and damp conditions. *Journal of Molluscan Studies* 2009; 75: 192-194.
- Watanuki N and Kawamura G. A review of cuttlefish basket trap fishery. *South Pacific Study*1999; 19: 31-48.

- Watanuki N, Hirayama I and Kawamura G. Why do cuttlefish *Sepia esculenta* enter basket
  traps? Space occupation habit hypothesis. *Fisheries science* 2000; 66: 190-197.
- 532

#### 533 Sepiolidae

- Hanlon RT, Claes MF, Ashcraft SE and Dunlap PV. Laboratory culture of the sepiolid squid *Euprymna scolopes*: a model system for bacteria-animal symbiosis. *The Biological Bulletin*1997; 192: 364-374.
- Jones N and McCarthy I. Aquaculture rearing techniques for the common cuttlefish *Sepia officinalis* and the Atlantic bobtail squid *Sepiola atlantica*. *Substainable Environmentally Friendly Diversification of Aquaculture for the Atlantic region of Europe*. 2009, pp.6-13.
- Swift K, Johnston D and Moltschaniwskyj N. The digestive gland of the southern dumpling
  squid (*Euprymna tasmanica*): structure and function. *Journal of experimental marine biology and ecology* 2005; 315: 177-186.
- 543
- 544 Squid
- 545 Budelmann BU. Cephalopoda. *The UFAW handbook on the care and management of laboratory and*546 *other research animals* 2010: 787-817.
- 547 Chabala LD, Morello RS, Busath D, Danko M, Smith-Maxwell CJ and Begenisich T. Capture,
  548 transport, and maintenance of live squid (*Loligo pealei*) for electrophysiological studies.
  549 *Pflügers Archiv* 1986; 407: 105-108.
- Hanlon RT. Maintenance, rearing, and culture of teuthoid and sepioid squids. *Squid as experimental animals*. Springer, 1990, pp.35-62.
- Hatfield EMC, Hanlon RT, Forsythe JW and Grist EPM. Laboratory testing of a growth
  hypothesis for juvenile squid *Loligo pealeii* (Cephalopoda: Loliginidae). *Canadian Journal of Fisheries and Aquatic Sciences* 2001; 58: 845-857.
- 555 Ikeda Y, Kidokoro H and Uji R. Notes on an exhausted Japanese common squid, *Todarodes*556 *pacificus* (Cephalopoda: Ommastrephidae), with an unusually short arm. *Bulletin of the*557 *College of Science University of the Ryukyus* 2004; 77: 143-148.
- Kaplan MB, Mooney TA, McCorkle DC and Cohen AL. Adverse effects of ocean acidification
  on early development of squid (*Doryteuthis pealeii*). *PLoS One* 2013; 8: e63714.
- LaRoe ET. The culture and maintenance of the loliginid squids *Sepioteuthis sepioidea* and
   *Doryteuthis plei. Marine Biology* 1971; 9: 9-25.
- 562 Matsumoto G. Transportation and maintenance of adult squid (*Doryteuthis bleekeri*) for
   563 physiological studies. *The Biological Bulletin* 1976; 150: 279-285.
- 564 O'Dor RK, Durward RD and Balch N. Maintenance and maturation of squid (*Illex illecebrosus*)
  565 in a 15 meter circular pool. *The Biological Bulletin* 1977; 153: 322-335.
- 566 Olmos-Pérez L, Pierce GJ, Roura Á and González ÁF. Barcoding and morphometry to identify
  567 and assess genetic population differentiation and size variability in loliginid squid
  568 paralarvae from NE Atlantic (Spain). *Marine Biology* 2018; 165: 136.
- 569 Otero J, Álvarez-Salgado XA, González ÁF, Souto C, Gilcoto M and Guerra Á. Wind-driven
  570 upwelling effects on cephalopod paralarvae: *Octopus vulgaris* and Loliginidae off the
- 571 Galician coast (NE Atlantic). *Progress in Oceanography* 2016; 141: 130-143.

- 572 Perretti CT, Zerofski PJ and Sedarat M. The spawning dynamics of California market squid
  573 (*Doryteuthis opalescens*) as revealed by laboratory observations. *Journal of Molluscan Studies*574 2016; 82: 37-42.
- Zakroff C, Mooney TA and Wirth C. Ocean acidification responses in paralarval squid
  swimming behavior using a novel 3D tracking system. *Hydrobiologia* 2018; 808: 83-106.
- 577
- 578 Octopus
- 579 Boyle PR. Methods for the aquarium maintenance of the common octopus of British waters,
  580 *Eledone cirrhosa. Laboratory Animals* 1981; 15: 327-331.
- 581 Budelmann BU. Cephalopoda. *The UFAW handbook on the care and management of laboratory and*582 *other research animals* 2010: 787-817.
- Castellano GC, da Veiga MPT, Mazzini FS, Vidal EAG and Freire CA. Paralarvae of *Octopus vulgaris* Type II are stenohaline conformers: relationship to field distribution and dispersal.
   *Hydrobiologia* 2018; 808: 71-82.
- Fuentes L, Iglesias J, Sánchez FJ, Otero JJ, Moxica C and Lago MJ. Métodos de transporte de paralarvas y adultos de pulpo *Octopus vulgaris* Cuvier, 1797. *Boletín Instituto Español de Oceanografía* 2011; 21: 155-162.
- Kawashima S, Amida A and Ikeda Y. Preliminary report of specific behaviours of juvenile
  greater blue-ringed octopus *Hapalochlaena lunulata* (Quoy and Gaimard, 1832). *Molluscan Research* 2019; 39: 291-295.
- 592 Shepherd B, Ross R and Avila M. Collection, Transport and Husbandry of the Coconut
- 593Octopus, Amphioctopus marginatus (Taki, 1964) from the Philippines. In: Williams GCGTM
- (ed) The Coral Triangle: The 2011 Hearst Philippine Biodiversity Expedition. California
- 595Academy of Sciences, San Francisco, California 94118, USA, 2014.